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# Herbicide resistance modelling: past, present and future

# Michael Renton,<sup>a\*</sup> Roberto Busi,<sup>a</sup> Paul Neve,<sup>b</sup> David Thornby<sup>c</sup> and Martin Vila-Aiub<sup>a,d</sup>

#### Abstract

Computer simulation modelling is an essential aid in building an integrated understanding of how different factors interact to affect the evolutionary and population dynamics of herbicide resistance, and thus in helping to predict and manage how agricultural systems will be affected. In this review, we first discuss why computer simulation modelling is such an important tool and framework for dealing with herbicide resistance. We then explain what questions related to herbicide resistance have been addressed to date using simulation modelling, and discuss the modelling approaches that have been used, focusing first on the earlier, more general approaches, and then on some newer, more innovative approaches. We then consider how these approaches could be further developed in the future, by drawing on modelling techniques that are already employed in other areas, such as individual-based and spatially explicit modelling approaches, as well as the possibility of better representing genetics, competition and economics, and finally the questions and issues of importance to herbicide resistance research and management that could be addressed using these new approaches are discussed. We conclude that it is necessary to proceed with caution when increasing the complexity of models by adding new details, but, with appropriate care, more detailed models will make it possible to integrate more current knowledge in order better to understand, predict and ultimately manage the evolution of herbicide resistance.

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Keywords: computer; simulation; evolution; integration; genetics

#### 1 INTRODUCTION

Herbicide resistance is a serious agricultural issue that threatens the sustainability of world food production.<sup>1-3</sup> Hundreds of biotypes of weeds resistant to commonly applied herbicides are now found throughout the world.<sup>4,5</sup> The emergence of herbicide resistance is an evolutionary and ecological process, influenced by many interacting factors; these include the chemistry and rate of herbicides applied, other non-chemical weed management, the particular ecological and biological traits of the weed species or ecotype being studied, environmental conditions and the possible biochemical and molecular mechanisms capable of conferring effective resistance, and their underlying genetic basis. Computer simulation modelling provides an important tool for helping to build an integrated understanding of how these different factors interact to affect the evolutionary and population dynamics of herbicide resistance, and thus helps in predicting and managing how agricultural systems will be affected.

In this review, we (1) expand on why computer simulation modelling is such an important tool and framework for understanding, predicting and managing herbicide resistance, (2) explain what questions related to herbicide resistance have been addressed to date using simulation modelling, (3) discuss the modelling approaches that have been used, focusing first on the earlier, more general approaches, and then on some newer, more 'cutting-edge' approaches, (4) consider how these approaches could be further developed in future, by drawing on modelling techniques that are already employed in other areas, and finally (5) discuss what new questions and issues could be addressed using these new

approaches. This review focuses on modelling the evolution of herbicide resistance, in contrast to other recent overviews that have taken a different or wider focus, such as modelling the evolution of resistance to pesticides and drugs in general, or modelling weed population dynamics in general. We also focus on models that represent or simulate the dynamic evolutionary and population processes underlying the evolution of resistance, while recognising the value of many other types of modelling, such as more empirical modelling of herbicide risk assessment or economic modelling that does not explicitly represent genetics.  $^{16-21}$ 

#### 2 USE AND VALUE OF MODELLING

Computer modelling provides a tool for integrating current knowledge and hypotheses regarding the different factors and processes that influence evolution of resistance (Fig. 1). This can help

- \* Correspondence to: Michael Renton, School of Plant Biology M090, The University of Western Australia, WA 6009, Australia. E-mail: michael.renton@uwa.edu.au
- a School of Plant Biology, Australian Herbicide Resistance Initiative and Institute of Agriculture, The University of Western Australia, Crawley, WA, Australia
- b School of Life Sciences, University of Warwick, Coventry, UK
- c Queensland Department of Agriculture, Fisheries and Forestry, Toowoomba, Qld, Australia
- d IFEVA-CONICET, Facultad de Agronomía, Universidad de Buenos Aires (UBA), Buenos Aires, Argentina



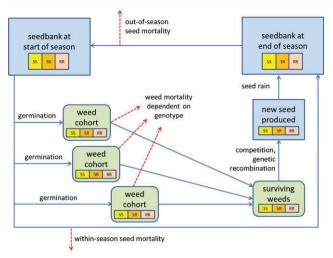


Figure 1. Conceptual model illustrating the typical dynamics simulated in a model of herbicide resistance evolution in an annual weed. The boxes usually represent the frequencies of different genotypes in the various weed and seedbank subpopulations and cohorts; sizes or densities of the subpopulations may also be represented (here, three genotypes are represented: a homozygous susceptible SS, a homozygous resistant RR and a heterozygote SR, under the assumption of a single gene, but any number is possible). A number of steps occurring within a year are simulated, resulting in transitions between the various weed and seedbank subpopulations and cohorts (solid arrows) or mortality (dashed arrows). At the start of the growing season, there exists a dormant seedbank consisting of seeds of different genotypes. One or more germination events occur. each resulting in a proportion of the seeds becoming a weed cohort (here, three germination events and cohorts are represented, but any number is possible). Herbicide application results in weed mortality, with a different mortality for different genotypes. Surviving weeds produce new seed, with the number of new seeds influenced by competition between weed cohorts and the crop, and the genotype of the new seeds determined by genetic recombination between the surviving weeds. This seed is added to the remaining dormant seedbank that has survived within-season seed mortality to form the end-of-season seedbank. Seed that survives out-of-season seed mortality then becomes the starting seedbank for the next season, and thus the simulation continues, with all these within-year processes simulated repeatedly to represent the population and evolutionary dynamics over several years. The process of evolution is driven by the differential survival of weeds under herbicide treatment, so that over a number of years the proportions and numbers of different genotypes will vary. Non-chemical weed treatments can be included in the model at almost any place; for example, harvest seed control techniques would result in a proportion of the seed produced being removed and not combined with the dormant seedbank, or higher crop sowing density might result in higher competition on the surviving weeds and thus lower seed production. Herbicide and non-chemical management can be assumed to be the same each year, or to vary from year to year. Variable environmental conditions may affect any of the processes in the model (proportion of germination for each cohort, competition, seed mortality, herbicide efficacy, etc.), or an 'average season' can be assumed in each year. Typically, the dynamics are simulated over several years, and the changing genotype frequencies and population densities are tracked until either a set number of years is reached or some other stopping criterion (such as weed population reaching a critical density threshold) is reached.

in understanding and predicting herbicide resistance evolution, and thus potentially in designing and evaluating effective proactive strategies to delay or even avoid its appearance, <sup>22,23</sup> rather than just reactively trying to manage populations that are already resistant. <sup>24</sup> Modelling allows and indeed encourages herbicide resistance to be approached from evolutionary and ecological perspectives likely to yield more complete understanding and, ultimately, better management outcomes. <sup>25,26</sup> Many important aspects of resistance to pesticides, such as the nature of underlying

mechanisms, their genetic bases and the levels of resistance they confer, can be best studied in laboratory, glasshouse or field experimentation. However, it is impossible to capture fully the long-term and large-scale dynamics of resistance evolution in such experimental systems. This is because the processes occur among millions or more usually billions of individuals, across huge areas and over many years. Experiments or trials on these scales are likely to be too expensive to be feasible, take too long to be useful and pose unjustifiable biosecurity risks by producing large numbers of resistant individuals, but computer simulation enables safe and quick virtual experiments to be conducted to investigate how various management options will affect the rate of emergence of herbicide resistance in large populations across large areas and long timeframes. Computer modelling can also represent aspects of the system that are very difficult to measure accurately in reality, such as the frequency of rare genes and the nature of soil seed banks, and the way these change with time. This can provide an insight into the way these hidden factors are likely to be affected by management factors, interact with ecological factors and affect herbicide efficacy and weed population sizes. Modelling also allows a much wider range of factors and interactions to be varied than would be possible in real experiments, giving a more complete understanding of the causal relationships determining the way in which resistance evolves.

It is also important to recognise the limitations of computer modelling as a tool for understanding and particularly for predicting herbicide resistance evolution. The accuracy of a simulation model's predictions depends on the accuracy of (1) the representation of underlying processes within the model, (2) the parameter values used in the model and (3) the definition of initial conditions.<sup>27</sup> Errors in model assumptions can lead to large errors in predictions.<sup>28</sup> For example, the rate at which herbicide resistance evolves will depend strongly on the initial frequency of resistance alleles within the population; this initial frequency may vary widely between different populations, and yet it is difficult accurately to measure this initial (or current) frequency in a given population. However, even if quantitatively accurate prediction of time to emergence of resistance is not possible because exact values for certain parameters are not known, sensitivity analysis of parameters with unknown values can be used to estimate their relative importance, thus identifying important data gaps and helping to prioritise further empirical research. Such sensitivity analysis may also reveal that relative effects do not depend on the unknown parameter values; for example, more frequent use of a herbicide may lead to faster evolution of resistance to that herbicide for a very wide range of initial frequencies of resistance alleles.

## 3 HERBICIDE RESISTANCE ISSUES ADDRESSED WITH MODELLING

#### 3.1 Time to resistance

One of the first answers many managers would like a herbicide resistance model to provide is how long it will take until they have a resistance problem. Ideally, the model would be able to predict how long it will take under the current weed management strategy, and, for comparison, how long it would take under possible alternative weed management strategies. Almost all herbicide resistance models provide a prediction of 'time to resistance', with some defining this in terms of a threshold frequency of resistance within the population<sup>29,30</sup> and others seeking a definition of more direct relevance to the farmer, such as time until weed populations reach a critical level, or until herbicide efficacy falls below a certain



threshold.<sup>31,32</sup> Modelling studies then typically look at how this 'time to resistance' depends on weed management, and other ecological, biological and genetic factors (Table 1). However it is defined, models have predicted that this 'time to resistance' is very sensitive to parameters that are almost impossible to measure in practice, and that may vary considerably between populations of the same species. In particular, the initial resistance allele frequency has a substantial effect on predicted time to resistance across a range of different models, and is likely to vary substantially in real populations – few of which remain entirely unselected in any case.<sup>29,33,34</sup> Therefore, much herbicide resistance modelling work has focused on predicting whether certain management options are likely to increase or decrease the time to resistance in relative terms, rather than absolute time estimates.

#### 3.2 Rotations and mixtures

One management option often recommended to delay the evolution of herbicide resistance is the use of herbicide rotations and mixtures/combinations, and this has been a popular topic for modelling as well (Table 1).30,32,35 'Combination' strategies, defined as strategies involving 'multiple intragenerational killing', can be considered to include both the application of physical mixtures of different herbicides and strategies such as 'double knockdown' that do not necessarily involve a literal mixing of different herbicides but still apply more than one herbicide within each generation.<sup>35</sup> Empirical studies have provided evidence that mixtures/combinations may be more effective than rotations in delaying resistance evolution through herbicide selection,<sup>36</sup> and modelling has supported this empirical evidence.  $^{34,35,37,38}$  The issue of mixtures is growing in importance, as new crop technologies with resistance to multiple herbicides are developed.<sup>39</sup> Positive cross-resistance, where resistance to one herbicide also confers some resistance to another, is likely greatly to reduce the usefulness of rotations, mixtures and multiple-herbicide-resistant crop technologies as control measures;<sup>40</sup> to date, this issue has sometimes been ignored in modelling studies. Modelling has predicted that the efficacy of rotation and mixtures will depend on the fitness costs associated with resistance and negative cross-resistance (where resistance to one herbicide reduces resistance to another) between the herbicides used. 32,41,42

#### 3.3 High and low dose

Another, somewhat controversial, management issue addressed with modelling has been the question of herbicide dose, or the 'dose rate debate' as it is sometimes called (Table 1). 26,43,44 Models have consistently predicted that higher selection pressure (usually associated with higher dose) leads to faster increases in the frequency of resistance alleles within a population over time when resistance is assumed to be conferred by a single dominant gene of major effect (monogenic resistance).<sup>29,45</sup> However, herbicide resistance modelling has suggested that relatively low doses are likely to lead to resistant weed problems faster when effective resistance is conferred by multiple genes acting together, and any one of these genes can only confer a partial minor level of resistance (polygenic resistance).<sup>31,46</sup> In this case, low doses tend to allow individuals with partial levels of resistance to survive, 47,48 which then cross with individuals with partial levels of resistance conferred by other genes, producing some progeny with more of these minor genes 'stacked together' and thus higher resistance. This means that, over a number of years, weed populations can evolve resistance more quickly under low doses; in combination

with the lower kill rates achieved with lower herbicide rates, this means that weed populations may increase in size considerably faster than if higher doses were used. This is one case where modelling indicates differences between resistance evolution in weeds and other organisms, with modelling for fungicide resistance indicating that low rates pose no risk of increased rates of evolution.<sup>43</sup> The modelling also provides an insight into why we might expect such differences might be expected between weeds and fundi, because sexual recombination is generally less frequent with fungi, and fungal populations do not generally need several years to build up populations. Modelling has also suggested that a 'revolving dose' strategy of alternating relatively high doses and relatively low doses may be effective when both polygenic and monogenic resistance genes are present at low levels in a population; the low doses reduce selection pressure for the monogenic resistance, and the high doses help to stop the accumulation of polygenic resistance alleles. 49,50 For organisms besides weeds, when it is possible to apply a pesticide dose for which the expression of resistance is recessive, modelling has predicted that a strategy combining areas of high-dose applications with areas of refuges (no dose) would be effective in delaying resistance evolution, 51-55 and it appears that the strategy has been at least somewhat effective,55 even if limitations of the modelling and strategy must be acknowledged.<sup>28,55</sup> However, similar strategies in weeds have not received much attention from modellers because of the difficulty of finding situations where the required combination of an economically viable herbicide dose and a recessive expression of resistance exists.

#### 3.4 Tillage

Soil cultivation has long been used as a weed control method, and occasional strategic tillage has been recommended for managing weeds in reduced-tillage systems,<sup>56,57</sup> along with harvest seed control measures.<sup>58–60</sup> These non-chemical control methods are promoted as ways to deal with populations that already have resistance or to delay resistance evolution by removing the need for some herbicide applications within the system. Modelling has predicted that the risk of resistance evolution can also be reduced by adding tillage to a system without removing any herbicide applications, by gaining additional weed kill through soil disturbance at sowing<sup>38,61</sup> or by burying resistant populations through occasional strategic soil inversion (Table 1).<sup>62</sup>

### 3.5 Interactions between management, genetics and ecology

As well as looking directly at the effect of management issues on rates of evolution of herbicide resistance rates of evolution, modelling has been used to investigate how other biological, ecological or genetic factors may influence rates of evolution, or even how biological or genetic factors may interact with management to affect rates of evolution (Table 1). Some of the factors that have been considered include the underlying genetic basis of resistance (inheritance), the mating system, gene flow, fecundity, seed dormancy and longevity of soil seed banks, and ecological fitness of different resistance genotypes.<sup>32-34,45,62-65</sup> For example, investigating interactions between seed dormancy and germination and the timing of when a herbicide is applied within a growing season in relation to other herbicides, and so which herbicides affect which weed cohorts, can help to explain different rates of evolution of resistance.34,66 Many of these ecological/biological factors apply specifically to weeds, as opposed to other organism types,



justifying the development of weed-specific models. Studies have also considered how biological or genetic factors may interact with management options to influence the rate at which populations might regress from evolved resistance and become more susceptible again, <sup>67,68</sup> and how these factors influence the ideal weed density thresholds that should be maintained in fields. <sup>69,70</sup>

#### 3.6 Spatial spread

Spatial spread of resistance through pollen and seed dispersal is another issue that has been addressed with modelling (Table 1). There has been some spatially explicit modelling of resistance evolution dynamics that has considered the spread both within and among fields.<sup>71–77</sup> Modelling of the spread of resistance

Table 1. Published studies that have used modelling to understand and/or predict the evolution of resistance to herbicides, ordered by publication year. The superscript next to each year is the number in the References list. For each study, the table shows whether the study explicitly tested the effect of certain management, genetic and biological/ecological factors on the evolutionary dynamics. It also shows whether the model used in the study had certain characteristics that would tend to increase model realism but at the expense of increased model complexity and/or reduced generality. Management factors include herbicide rotation, herbicide mixture, mosaic, i.e. spatial heterogeneity in herbicide application, herbicide dosage, tillage/cultivation or other management options. Genetic factors include the initial frequency of resistance alleles, the mutation frequency, dominance, the strength of resistance, the possibility of cross-resistance, the number of genes involved in resistance and epistasis. Biological/ecological factors include fitness penalties for resistance alleles, seed dormancy and/or longevity, dispersal and/or gene flow, outcrossing rates and initial population size. Model characteristics include whether the model represents population dynamics, as opposed to just genetic dynamics, density dependence, spatial heterogeneity and multiple genes/loci, and also whether the model is stochastic, as opposed to deterministic, whether it was simulated on a computer to obtain results, as opposed to being solved analytically, and whether it was explicitly constructed to represent a specific weed species

Year	Management factors						Genetic factors							Biological/ecological factors						Model characteristics						
	rotation	mixture	mosaic	dosage	tillage	other	initial frequency	mutation	dominance	strength	cross-resistance	number genes	epistasis	fitness	dormancy	dispersal	outcrossing	init pop size	pop dynamics	density depend.	spatial heterog.	multiple genes	stochastic	simulated	specific species	
1978 <sup>45</sup>							у	у						у	у											
1990 <sup>32</sup>	у						y	,						у	у											
1990 <sup>68</sup>	,			у			,							у	,	у	у		у					у		
1992 <sup>63</sup>	у			,										у	у	,	,		y					,	у	
1992 <sup>70</sup>	,					у								,	,	у			,		у			у	y	
1994 <sup>64</sup>	у	у				,	у							у		,					,			у	,	
1996 <sup>33</sup>	у	,					у	у	у					,										,		
1996 <sup>90</sup>	,			у		у	у	,	,							у			у	у	у				у	
1998 <sup>49</sup>	у			у		,	,		у							,			у	y	,	у		у	,	
2000 <sup>93</sup>	y			,	у		у		,									у	ý	ý		,	у	у	у	
2001 <sup>29</sup>	у	у		у	ĺ		у		у			у							ý	ý		у	,	у	y	
2001 <sup>94</sup>	у			Í	у		•	у									у	у	у	у			у	у	у	
2002 <sup>74</sup>	у		у	у				•							У	у	Í		y	y	у			у	,	
2002 <sup>92</sup>	у				у														у					у	у	
2003 <sup>37</sup>	у	у															у	у	у	У		у	у	у		
2003 <sup>66</sup>		у				у	у		у					у	у				у	у		у	у	у	У	
2003 <sup>38</sup>	у	у			у	у	у	У							у			у	У	у		у	у	у	у	
2004 <sup>73</sup>																У			У	У	У			У		
2006 <sup>86</sup>																										
2007 <sup>77</sup>	У		У						У					у			у		У	У	У			У		
2008 <sup>30</sup>	У	у								у									У					У	У	
2008 <sup>34</sup>	У	у		У		у	у		У					У	У		У	у	У	У			у			
2008 <sup>72</sup>	У																				У					
2008 <sup>76</sup>	У		У						У					У			У		У	У	У			У		
2009 <sup>42</sup>		у							У	У	У			У			У		У	У				У		
2009 <sup>87</sup>	У			У	У	У													У				У	У	У	
2011 <sup>31</sup>				У					У	У		У	У						У	У		У	У	У		
2011 <sup>88</sup>		У					У								У			У	У	У			У	У	У	
2011 <sup>89</sup>	У	У			У														У	У			У	У	У	
2012 <sup>46</sup>				У					У	У		У	У						У	У		У	У	У	У	
2012 <sup>62</sup>	У			У	У							У					У		У	У		У	У	У		
2013 <sup>61</sup>	У	У		У	У	У									У		У		У				У	У	У	
2013 <sup>91</sup>	У	У			У	У	У								У			У	У	У			У	У	У	
2013 <sup>100</sup> 2013 <sup>104</sup>												У		У					У	У		У		У		
2013104	У			У	У							У				У			У	У	У	У	У	У		



genes from transgenic crops has also received considerable attention,<sup>78–80</sup> and this modelling has potential to inform the modelling of the 'natural' emergence of resistance in weeds under selection from herbicide treatments in the way that it represents the flow of genes through pollen and possibly seed dispersal.

### 4 APPROACHES TO MODELLING THE EVOLUTION OF RESISTANCE

#### 4.1 Simpler approaches

Early approaches to modelling the dynamic evolution of resistance usually aimed to capture the most important factors involved in relatively simple ways (Table 1). Simpler models also tend to be relatively general and abstract, and so they did not necessarily contain features that were specific to herbicides and weeds, but could also be thought of as representing other types of organism, although a few models were specifically developed to represent weeds.<sup>32,45,68,81,82</sup> In such models, the basis of resistance was usually assumed to be monogenic, and the gene was often assumed to be fully dominant, leading to an assumption of just two phenotypes - one fully susceptible and one completely resistant. These models were usually based on population density, in that their state variables represented the frequency or proportions of different alleles, genotypes or phenotypes in the population; they thus predicted when resistance might appear, but not what densities of resistant individuals to expect. These frequencies could usually take any value between 0 and 1, meaning that the discrete nature of real populations and genomes was not accounted for. These models did not typically represent seasonal variation effects or ecological aspects specific to weeds, such as dormant seed banks persisting across seasons or staggered germination within a season resulting in a number of different weed cohorts.

#### 4.2 Adding more details of the real biological system

With time, models of herbicide resistance evolution began to include representation of more aspects of the real biological system (Table 1). In many cases this involved making the model more specific to weeds and herbicides (rather than general organisms and pesticides), and in some cases specific to particular weed species or agricultural systems (see below for examples). In other cases, genetic details were added without ecological/biological details that made the model specific to weeds.<sup>49,83</sup>

In the simplest approaches, actual weed population density is largely ignored, and only the frequency of the susceptible/resistant genotypes is represented. However, in reality it is weed population sizes or densities that concern farmers, and modellers have recognised this and included representation of plant and seed numbers, along with genotype frequencies (e.g. Renton et al.<sup>31</sup> and Gardner et al.<sup>49</sup>). With population densities comes the need to consider density-dependent nature of seed production, self-thinning and mortality, and intra- and interspecific resource competition. These have often been represented in fairly simple descriptive ways, such as using a hyperbolic competition function like that developed by Firbank and Watkinson, 84,85 where seed production per unit area tends towards a fixed maximum at high densities (see Renton et al.<sup>31</sup> and Diggle et al.<sup>37</sup> for examples), although more recently resistance evolution models have been linked with more complex crop growth simulation models that handle the simulation of crop-weed competition with much greater temporal and spatial resolution and biological realism. 61,86,87

As mentioned, some herbicide resistance models have started to focus guite specifically on the details of particular species, herbicides and systems. Such modelling may, for example, focus very specifically on glyphosate resistance evolution in Palmer amaranth (Amaranthus palmeri) in glyphosate-resistant transgenic cotton in the southern United States<sup>88,89</sup> or glyphosate resistance evolution in awnless barnyard grass (Echinochloa colona) in glyphosate-resistant transgenic cotton in north-east Australia. 61 Herbicide resistance modelling has been applied specifically to parasitic weeds<sup>90</sup> and to one particular species and herbicide, <sup>91–94</sup> and models have been based on field data for the dynamics and relative fitness of resistant and susceptible biotypes of a particular species, making them even more specific to the context in which those data were collected.<sup>63,81</sup> In some cases, this specific focus has been achieved by linking evolutionary models with crop models, meaning that predictions from individual runs are specific not only to the crops and weed species but also to the soil type and historical weather of a particular field.61,87

As mentioned above, most earlier models of herbicide resistance evolution assumed a monogenic basis for resistance, following the fact that, for most early studied cases of resistance, when a genetic basis was identified, it was monogenic. However, there is a growing concern regarding polygenic resistance, which is more associated with problematic cross-resistance to multiple unrelated types of herbicide, 95-97 and for which selection patterns may be different from selection of monogenic resistance.<sup>31</sup> When earlier resistance modellers tackled the issue of polygenic resistance, they tended to use a quantitative genetics approach, based on several assumptions, including that resistance is conferred by a large number of unlinked genes, each of very small additive effect.98 More recently, modellers have represented multiple genes and their interactions more explicitly, allowing a more detailed and realistic representation of a polygenic basis for herbicide resistance, 31,99,100 or other pesticides. 83,101,102 This explicit approach also allows for representation of the linkage disequilibrium between alleles that is likely to emerge when very strong selection pressures are driving evolutionary processes, 83 but that is ignored in the 'traditional' quantitative genetics approaches. Representing the genetics more explicitly has the advantage that initial allele frequencies, mutation frequencies and fitness penalties can also be represented separately for each resistance gene and allele; it also means that values for these parameters need to be defined and justified.

Spatial heterogeneity is another aspect of biological reality that has been added to some herbicide resistance models in more recent times. When modelling herbicide resistance, spatial heterogeneity in weed and seed densities is likely to be important, as is spatial heterogeneity in the frequencies of resistance alleles. To represent spatial heterogeneity, resistance models can take a relatively simple and implicit approach of just adding variation to parameters likely to vary across space, such as dose, 31,101,103 or they can represent space and variation in population density and genetic frequencies more explicitly. Such spatially explicit models can represent variation in space in one dimension, along a transect or gradient, or down through depths in a soil profile, 62 in two dimensions across a field or a landscape of many fields 71 – 77,104 or potentially in all three dimensions.

Uncertainty and stochasticity are also important aspects of herbicide resistance evolution. While earlier models tended to be deterministic, more recent models have included random stochastic processes to represent some of the uncertainty observed in real systems. For example, when weeds are sprayed with herbicide, a model can simulate that exactly 95% of susceptible plants will die



(deterministic), or that each plant in the population has a 95% chance of dying (stochastic). Stochasticity is important in resistance evolution, because it often depends on relatively rare occurrences (gene mutations, the chance survival of an intermediate resistant genotype, sexual reproduction resulting in two particular complementary resistant genes coming together in one organism to create a new strongly resistant genotype). Stochasticity can also represent sources of variation that are not explicitly modelled, such as plants having different sizes, and thus different chances of being killed by a herbicide, even if having the same resistance genes. Another source of uncertainty is basic uncertainty about how well model parameter values represent the real system. Understanding the uncertainty in predictions introduced by stochastic processes within the model and/or uncertainty in model parameter values generally requires multiple simulation runs to produce a distribution of results that represent the full range of likely outcomes (Monte Carlo simulation).88 Another possibility is a sensitivity analysis that looks at the effect of varying each model parameter one at a time.31,45

Another approach that has been used in herbicide resistance modelling to some extent is individual-based modelling (IBM), as opposed to population density approaches. IBM involves explicit representation of the individuals within a population, along with their individual characteristics. <sup>105 – 108</sup> IBM approaches have been used in modelling resistance in a range of specific types of organism, including insects and parasitic nematodes, <sup>53,54,83,101,109,110</sup> and in more theoretical non-organism-specific studies, <sup>111</sup> but only to a limited extent with weeds. <sup>31</sup>

#### 5 THE ROAD AHEAD

#### 5.1 Individual-based modelling approaches

There are many open avenues for methodological development in herbicide resistance modelling, and many applications that could be addressed with these new models. IBM approaches are promising for a deeper understanding of resistance evolution and the complex interactions involved because they can incorporate more of the biological detail and variability of real systems. 106 This can represent more of the biological variation within populations, and particularly how various ecological and genetic traits of individuals may become correlated or dependent over time under certain management regimes. This would allow, for example, a better understanding of the complex interrelationships between genetics and ecological 'fitness penalties' operating at different points in the lifecycles of plants, such as reduced seed and/or pollen production, increased mortality or diminished competitiveness in certain conditions.<sup>112</sup> As another example, IBM could help to unravel some of the complex interactions between evolution of seed dormancy and germination traits, evolution of chemical herbicide resistance and timing of herbicide applications, and thus help in understanding observed relationships between seed dormancy and herbicide resistance. 113-116 To date, biological parameters in models, such as those controlling dormancy, competitiveness or flowering time and thus fecundity, have usually been assumed to be fixed, whereas in reality they are likely to change over generations under different simulated agronomic conditions, just like chemical resistance; IBM can help to account for this coevolution. One caution for IBM approaches is that the additional realism and better representation of biological complexity usually come at the cost of additional model complexity, which may make models more difficult to calibrate, verify and analyse.105,106

#### 5.2 Spatially explicit models

Making herbicide resistance models more spatially explicit may also be important. The assumptions that weed populations at the scale of a field exhibit spatially homogeneous densities and genetic frequencies, and random mating, are clearly not realistic. It has already been shown that the finite nature of weed populations can have real implications for model predictions, even when modelled populations are large and not spatially differentiated;<sup>37</sup> it is even more likely to be important when we consider that real weed populations of some species may be quite patchy and have low seed dispersal. This means that the possibility for local extinction of resistance alleles will be much higher than if the population is considered to be large and spatially homogenous. Spatially explicit herbicide resistance modelling should learn from the rich field of spatially explicit modelling of dispersal and population dynamics that has already been employed in a diverse range of fields, including invasion biology, pest management, gene flow from transgenic crops and conservation. 79,80,117-122 Fortunately, recent empirical work is already providing valuable information on the dispersal of pollen and seeds that will help to inform these models.  $^{123-126}$  Spanor tially explicit herbicide resistance models would make it possible to simulate, evaluate and design spatially targeted weed control options, such as managing resistant patches with different herbicides using targeted detection and spray technologies 127-131 or autonomous robotic vehicles. 132 They would also make it possible to understand the importance of seed and pollen dispersal in the spread of herbicide resistance, and help to predict the efficacy of management options based around limiting dispersal, such as limiting seed movement through farm equipment hygiene and crop seed screening, 133 or possibly limiting pollen flow by using barrier zones or spatial planning of land use accounting for dominant winds.76

#### 5.3 Competition

It may be worth considering the level of detail at which competition for resources between plants is represented within herbicide resistance models. Plant competition, and crop-weed competition in particular, has been modelled at varying levels of biological and dynamic realism, 134 including with crop growth approaches, 135 spatially explicit individual-based approaches 136 and functional-structural approaches that represent the dynamic above- and/or below-ground architecture of competing plants in high detail.<sup>137,138</sup> While these approaches may be overly complex for direct inclusion in evolutionary models that represent dynamics over many years, they might still have a role for informing the representation of competition in evolutionary models, and accounting for self-thinning and compensatory growth after herbicide mortality. A related issue is the representation of environmental and weather effects in herbicide resistance models. As mentioned, when a crop model is used to simulate competition and predict seed set, the seasonal effects of specific rainfall and temperature patterns on mortality, competition and seed set can be predicted, and their influence on long-term evolutionary dynamics can be accounted for.87 Effects of seasonal variation on seed germination timing and herbicide application efficacy may also be important, and incorporating these within herbicide resistance models should be considered.

#### 5.4 Genetics

The representation of genetic control of resistance could be further developed. For example, resistance models have not explicitly represented the processes involved in gene amplification,



which has been shown to be an important genetic mechanism underlying resistance evolution. 49,97,139 Herbicide resistance models with more explicit representation of relatively complex polygenic genetic mechanisms exist, but to date accurate data to parameterise these models have been lacking, and so the models have mostly been used to understand how different genetics would influence the efficacy of different management options.<sup>31</sup> However, new genomic tools and high-throughput techniques may help to provide such information, 140,141 and there are already examples for organisms besides weeds where such information on polygenic resistance has been obtained and incorporated into models. 101,102,142 Simulation models could also be used for 'inverse modelling', working backwards from observed data regarding mortality over several generations of herbicide selection in the field or glasshouse, to identify the likely genetic bases of any observed increase in resistance, and give insights into the evolutionary history of how the resistance is likely to have evolved. More realistic representation of resistance genetics would make it possible explicitly to represent fitness penalties related to individual genes and alleles, mutation rates at different loci and possibly the influence of environmental stresses and low herbicide doses on resistance mutation frequencies and selection efficiency. 143

#### 5.5 Economics

Economic considerations have not been much addressed in herbicide resistance evolutionary modelling, or indeed in most modelling of resistance evolution.<sup>6</sup> Agricultural economists have employed relatively simple models of weed population dynamics to address important economic questions regarding weed management, but these have generally ignored herbicide resistance, compared analyses with and without resistance or perhaps used very simple models of resistance evolution that assume a given herbicide will lose efficacy after a certain number of applications.<sup>16-21</sup> Similarly, weed ecologists' incorporation of economics into their model frameworks has usually been of the simplest sort, analysing only basic considerations of cost. The challenge of incorporating economic evaluation with greater biological detail so that the economic costs and benefits of different resistance management options can be properly evaluated still largely remains.

### 5.6 Advantages and disadvantages of greater detail and realism

Incorporating greater biological detail and reality into models will have benefits. Developing new herbicide resistance models with more explicit, detailed and realistic representation of space, variability between individual, competition, effects of environment and weather and genetic mechanisms will make it possible to revisit old questions previously addressed with models. Questions regarding herbicide rotation, mixtures, combinations, doses and tillage can be revisited to see if previous results hold up when processes are represented more realistically. We can also look more closely at how results depend on a full range of possibly influential factors, and interpret results in more meaningful ways. For example, models may provide a certain prediction when the genetic basis is assumed to be monogenic and dominant, the population is assumed to be homogeneous, pollen and seed dispersal are assumed to be infinite and mating is assumed to be perfectly random. But will the prediction be the same if the genetic basis is assumed to be polygenic and recessive, the population spatiality is assumed to be heterogeneous and limited dispersal

of pollen and seed and pollen competition are accounted for? And even if resistance evolves faster in terms of allele frequencies, does this mean that the farmer experiences faster growth in weed populations and quicker declines in yield and profit? New models will also make it possible to address new questions, such as the importance of gene flow through pollen or seed dispersal, or the feasibility of the spatially targeted resistant patch management mentioned previously. More detailed models will make it possible to integrate more current knowledge, from genetic and physiological mechanisms within cells to ecological processes of individual plants, populations and communities within fields, in order better to understand and predict and ultimately manage evolutionary processes operating across regions and many decades, including selection, migration, mutation and genetic drift.<sup>6</sup>

Incorporating greater biological detail and reality into models will also have costs. Building, calibrating, verifying, validating and analysing models costs time and money, and generally, the more detailed a model is, the more complex it will be, and the more time and money it will cost. There is also a direct cost of complexity; large complex models with many parameters are less transparent in the sense that it is less clear exactly how each parameter interacts with every other parameter to influence model outputs. This makes a complete analysis of a complex model difficult and eventually impossible, which is of course true of the real-world systems as well. For example, if the simplifying assumption is made that resistance is conferred by a single dominant gene, the model will include many fewer genetic parameters than if multiple genes are represented explicitly and separately, and different genes are allowed to have different effects and dominance and to interact with varying degrees of synergism or antagonism. In the latter case, much more testing of the effect of varying parameter values separately and in combination will be required than in the former case. Another issue is that, while simpler models can often be solved analytically to provide a complete solution,<sup>45</sup> more complex models usually need to be computationally simulated to obtain predictions or analyse the effect of varying parameter values (Table 1). Therefore, virtual experiments must be carefully planned to address questions and interactions of most interest and concern; while time consuming and providing less complete analysis than an analytical solution, these are still likely to be much faster, cheaper and safer than real-world experiments designed to address the same questions to the same level of generality. Nonetheless, an infinite number of models can be constructed, and an endless series of virtual experiments conducted, and so part of the ongoing goal must be to work out what details and differences really matter and where simplifications and generalisations can be made. For example, if we represent dynamic plant growth and competition processes at detailed scales of hours and individual leaves, do we obtain importantly different results to those obtained when plant competition is represented more simply and empirically?<sup>65</sup> If we model the spatial variation in weed density and allele frequency across a field in explicit detail at the scale of a square metre, do we achieve significantly different predictions or greater insights than assuming the whole field to be a homogenous mixed population? Does the model we develop to represent evolution of resistance to a specific herbicide in a particular weed species in a specific cropping system in a particular region of a certain country provide important new insights compared with a similar model developed for a different species or herbicide or system or region? We must seek a balance between generality and specificity, understanding that the strength of modelling as opposed to empirical study is more in



the direction of abstraction and generality. Rather than building a new, very specific model for every new situation, we can use a more general model to investigate how characteristics of different species, herbicides, resistance genetics and agricultural systems are likely to interact with different management options to affect the evolution of resistance. If modelling predicts that a certain management option is likely to be effective, even as weed competitiveness, seed production, outcrossing levels and dormancy traits are varied across a wide range of values, then we may conclude that there is little need to empirically identify those species traits. Or if modelling predicts that the efficacy of another management option is likely to depend strongly on the genetic basis of resistance, being highly effective for monogenic resistance but non-effective for polygenic, then we could focus research effort on better ways to identify genetic bases of resistance. A similar balance must be sought when looking at similarities and differences between organisms of different kingdoms; modellers of herbicide resistance should learn all they can from modellers of resistance evolution in insects, fungi and bacteria, 6,144,145 while also recognising that there are likely to be important differences and trying to understand what these are and what effect they have. In this way, modelling will continue to evolve and improve as a tool for integrating current knowledge and hypotheses regarding the different factors and processes that influence evolution of resistance, which will in turn help in understanding and predicting herbicide resistance better, and thus in designing and evaluating effective strategies to manage, delay or even avoid its appearance.

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#### REFERENCES

- 1 Mortensen DA, Egan JF, Maxwell BD, Ryan MR and Smith RG, Navigating a critical juncture for sustainable weed management. *Bioscience* **62**(1):75–84 (2012).
- 2 Powles SB and Yu Q, Evolution in action: plants resistant to herbicides. Annu Rev Plant Biol 61:317 – 347 (2010).
- 3 Powles SB and Shaner DL (eds), Herbicide Resistance and World Grains. CRC Press, Boca Raton, FL (2001).
- 4 Heap I, The International Survey of Herbicide Resistant Weeds. [Online]. Available: www.weedscience.com [24 September 2013].
- 5 Duke SO and Powles SB, Glyphosate-resistant weeds and crops. Pest Manag Sci 64(4):317–318 (2008).
- 6 REX Consortium, The skill and style to model the evolution of resistance to pesticides and drugs. Evol Applic 3(4):375–390 (2010).
- 7 Holst N, Rasmussen IA and Bastiaans L, Field weed population dynamics: a review of model approaches and applications. Weed Res 47(1):1–14 (2007).
- 8 Colbach N and Debaeke P, Integrating crop management and crop rotation effects into models of weed population dynamics: a review. Weed Sci 46:717–728 (1998).
- 9 Doyle CJ, Mathematical models in weed management. *Crop Prot* **10**(6):432–444 (1991).
- 10 Doyle CJ, A review of the use of models of weed control in Integrated Crop Protection. Agric Ecosyst Environ 64(2):165 – 172 (1997).
- 11 Renton M, Peltzer S and Diggle AJ, Understanding, predicting and managing weed seedbanks in agricultural systems with the Weed Seed Wizard, in *Proceedings of the 16th Australian Weeds Conference*, ed. by van Klinken RD, Osten VA, Panetta FD and Scanlan JC. Queensland Weeds Society, Brisbane, Qld, Australia (2008).
- 12 Parsons DJ, Benjamin LR, Clarke J, Ginsburg D, Mayes A, Milne AE et al., Weed Manager – a model-based decision support system for weed management in arable crops. Comput Electron Agric 65:155–167 (2009).

- 13 Stanton RA, Pratley JE, Hudson D and Dill GM, A risk calculator for glyphosate resistance in *Lolium rigidum* (Gaud.). *Pest Manag Sci* 64(4):402–408 (2008).
- 14 Werth J, Thornby D and Walker S, Assessing weeds at risk of evolving glyphosate resistance in Australian sub-tropical glyphosate-resistant cotton systems. Crop Past Sci 62(11):1002 – 1009 (2012).
- 15 Ferraro DO and Ghersa CM, Fuzzy assessment of herbicide resistance risk: glyphosate-resistant johnsongrass, Sorghum halepense (L.) Pers., in Argentina's croplands. Crop Prot 51:32–39 (2013).
- 16 Gorddard RJ, Pannell DJ and Hertzler G, An optimal control model for integrated weed management under herbicide resistance. Aust J Agric Resour Econ 39(1):71–87 (1995).
- 17 Gorddard RJ, Pannell DJ and Hertzler G, Economic evaluation of strategies for management of herbicide resistance. *Agric Syst* 51(3):281–298 (1996).
- 18 Monjardino M, Pannell DJ and Powles SB, Multispecies resistance and integrated management: a bioeconomic model for integrated management of rigid ryegrass (*Lolium rigidum*) and wild radish (*Raphanus raphanistrum*). Weed Sci 51(5):798–809 (2003).
- 19 Pannell DJ, Stewart V, Bennett A, Monjardino M, Schmidt C and Powles SB, RIM: a bioeconomic model for integrated weed management of *Lolium rigidum* in Western Australia. *Agric Syst* **79**(3):305–325 (2004).
- 20 Weersink A, Llewellyn RS and Pannell DJ, Economics of pre-emptive management to avoid weed resistance to glyphosate in Australia. Crop Prot 24(7):659–665 (2005).
- 21 Lawes R and Renton M, The Land Use Sequence Optimiser (LUSO): a theoretical framework for analysing crop sequences in response to nitrogen, disease and weed populations. *Crop Past Sci* **61**(10):835–843 (2010).
- 22 Norsworthy JK, Ward SM, Shaw DR, Llewellyn RS, Nichols RL, Webster TM *et al.*, Reducing the risks of herbicide resistance: best management practices and recommendations. *Weed Sci* **60**(Suppl. 1):31–62 (2012).
- 23 Friesen LJS, Ferguson GM and Hall JC, Management strategies for attenuating herbicide resistance: untoward consequences of their promotion. Crop Prot 19(8–10):891–895 (2000).
- 24 Beckie HJ, Herbicide-resistant weeds: management tactics and practices. *Weed Technol* **20**(3):793–814 (2006).
- 25 Neve P, Vila-Aiub M and Roux F, Evolutionary-thinking in agricultural weed management. New Phytol 184(4):783 – 793 (2009).
- 26 Neve P, Busi R, Renton M and Vila-Aiub MM, Evolutionary perspective on herbicide resistance (TBC). Pest Manag Sci this special issue (2014).
- 27 Haefner JW, Modeling Biological Systems: Principles and Applications. Springer, New York, NY (2005).
- 28 Gressel J, Problems in qualifying and quantifying assumptions in plant protection models: resultant simulations can be mistaken by a factor of million. Crop Prot 24(11):1007 – 1015 (2005).
- 29 Diggle AJ and Neve P, The population dynamics and genetics of herbicide resistance a modeling approach, in *Herbicide Resistance and World Grains*, ed. by Powles SB and Shaner DL. CRC Press, Baton Rouge, FL, pp. 61–99 (2001).
- 30 Gustafson DI, Sustainable use of glyphosate in North American cropping systems. Pest Manag Sci 64(4):409–416 (2008).
- 31 Renton M, Diggle A, Manalil S and Powles S, Does cutting herbicide rates threaten the sustainability of weed management in cropping systems? J Theor Biol 283(1):14–27 (2011).
- 32 Gressel J and Segel LA, Modelling the effectiveness of herbicide rotations and mixtures as strategies to delay or preclude resistance. *Weed Technol* **4**:186–198 (1990).
- 33 Jasieniuk M, Brûlé-Babel AL and Morrison IN, The evolution and genetics of herbicide resistance in weeds. Weed Sci 44:176–193 (1996).
- 34 Neve P, Simulation modelling to understand the evolution and management of glyphosate resistance in weeds. *Pest Manag Sci* **64**(4):392–401 (2008).
- 35 REX Consortium, Heterogeneity of selection and the evolution of resistance. Trends Ecol Evol 28(2):110–118 (2013).
- 36 Beckie HJ and Reboud X, Selecting for weed resistance: herbicide rotation and mixture. *Weed Technol* **23**(3):363–370 (2009).
- 37 Diggle AJ, Neve PB and Smith FP, Herbicides used in combination can reduce the probability of herbicide resistance in finite weed populations. Weed Res 43(5):371 – 382 (2003).



- 38 Neve P, Diggle AJ, Smith FP and Powles SB, Simulating evolution of glyphosate resistance in *Lolium rigidum* II: past, present and future glyphosate use in Australian cropping. *Weed Res* 43(6):418–427 (2003).
- 39 Green JM, Review of glyphosate and ALS-inhibiting herbicide crop resistance and resistant weed management. Weed Technol 21(2):547–558 (2007).
- 40 Beckie HJ and Tardif FJ, Herbicide cross resistance in weeds. *Crop Prot* **35**:15–28 (2012).
- 41 Gressel J and Segel LA, Negative cross resistance; a possible key to atrazine resistance management. Z Naturforsch C Biosci 45(5):470–473 (1990).
- 42 Jacquemin B, Gasquez J and Reboud X, Modelling binary mixtures of herbicides in populations resistant to one of the components: evaluation for resistance management. *Pest Manag Sci* 65(2):113–121 (2009).
- 43 van den Bosch F, Paveley N, Shaw M, Hobbelen P and Oliver R, The dose rate debate: does the risk of fungicide resistance increase or decrease with dose? *Plant Pathol* **60**(4):597 – 606 (2011).
- 44 Neve P, Challenges for herbicide resistance evolution and management: 50 years after Harper. Weed Res 47(5):365–369 (2007).
- 45 Gressel J and Segel LA, The paucity of plants evolving genetic resistance to herbicides: possible reasons and implications. *J Theor Biol* 75(3):349–371 (1978).
- 46 Manalil S, Renton M, Diggle A, Busi R and Powles SB, Simulation modelling identifies polygenic basis of herbicide resistance in a weed population and predicts rapid evolution of herbicide resistance at low herbicide rates. Crop Prot 40:114–120 (2012).
- 47 Neve P and Powles S, High survival frequencies at low herbicide use rates in populations of *Lolium rigidum* result in rapid evolution of herbicide resistance. *Heredity* 95(6):485–492 (2005).
- 48 Manalil S, Busi R, Renton M and Powles SB, Rapid evolution of herbicide resistance by low herbicide dosages. Weed Sci 59(2):210–217 (2011)
- 49 Gardner SN, Gressel J and Mangel M, A revolving dose strategy to delay the evolution of both quantitative vs major monogene resistances to pesticides and drugs. *Int J Pest Manag* 44(3):161–180 (1998).
- 50 Gardner SN, Agrawal AA, Gressel J and Mangel M, Strategies to delay the evolution of resistance in pests: dose rotations and induced plant defenses. Aspects Appl Biol 53:189 – 196 (1999).
- 51 Gould F, Potential and problems with high-dose strategies for pesticidal engineered crops. Biocontrol Sci Technol 4(4):451–461 (1994).
- 52 Gould F, Sustainability of transgenic insecticidal cultivars: integrating pest genetics and ecology. *Annu Rev Entomol* 43(1):701–726 (1998).
- 53 Storer NP, A spatially explicit model simulating western corn rootworm (Coleoptera: Chrysomelidae) adaptation to insect-resistant maize. J Econ Entomol 96(5):1530 – 1547 (2003).
- 54 Storer NP, Peck SL, Gould F, Van Duyn JW and Kennedy GG, Spatial processes in the evolution of resistance in *Helicoverpa zea* (Lepidoptera: Noctuidae) to *Bt* transgenic corn and cotton in a mixed agroecosystem: a biology-rich stochastic simulation model. *J Econ Entomol* **96**(1):156–172 (2003).
- 55 Bates SL, Zhao J-Z, Roush RT and Shelton AM, Insect resistance management in GM crops: past, present and future. *Nat Biotechnol* **23**(1):57–62 (2005).
- 56 Douglas A and Peltzer SC, Managing herbicide resistant annual ryegrass (Lolium rigidum Gaud.) in no-till systems in Western Australia using occasional inversion ploughing, in Proceedings of the 14th Australian Weeds Conference, ed. by Sindel BM and Johnson SB. Weed Society of New South Wales, Sydney, NSW, Australia, pp. 6–9 (2004).
- 57 Derksen DA, Anderson RL, Blackshaw RE and Maxwell B, Weed dynamics and management strategies for cropping systems in the northern Great Plains. *Agron J* **94**(2):174–185 (2002).
- 58 Walsh MJ and Powles SB, Management strategies for herbicide-resistant weed populations in Australian dryland crop production systems. Weed Technol 21(2):332–338 (2007).
- 59 Walsh M, Newman P and Powles S, Targeting weed seeds in-crop: a new weed control paradigm for global agriculture. *Weed Technol* **27**(3):431–436 (2013).
- 60 Walsh MJ, Harrington RB and Powles SB, Harrington Seed Destructor: a new nonchemical weed control tool for global grain crops. *Crop Sci* 52(3):1343–1347 (2012).

- 61 Thornby D, Werth J and Walker S, Managing glyphosate resistance in Australian cotton farming: modelling shows how to delay evolution and maintain long-term population control. *Crop Past Sci* 64(8):780–790 (2013).
- 62 Renton M, Flower K, Busi R, Thornby D, Airey M, Manalil S *et al.*, Modelling the effects of ecology, management and genetics on the evolution of herbicide resistance with PERTH, in *Developing Solutions to Evolving Weed Problems 18th Australasian Weeds Conference*, ed. by Eldershaw V. Weed Science Society of Victoria, Melbourne, Vic., Australia, pp. 275–278 (2012).
- 63 Mortimer AM, Ulf-Hansen PF and Putwain PD, Modelling herbicide resistance – a study of ecological fitness, in *Resistance '91: Achieve-ments and Developments in Combating Pesticide Resistance*, ed. by Denholm I, Devonshire AL and Hollomon DW. Springer, Dordrecht, The Netherlands, pp. 148–164 (1992).
- 64 Jasieniuk M and Maxwell BD, Populations genetics and the evolution of herbicide resistance in weeds. *Phytoprotection* **75**(4):25–35 (1994)
- 65 Renton M, How much detail and accuracy is required in plant growth sub-models to address questions about optimal management strategies in agricultural systems? AoB Plants 2011:plr006 (2011).
- 66 Neve P, Diggle AJ, Smith FP and Powles SB, Simulating evolution of glyphosate resistance in *Lolium rigidum* I: population biology of a rare resistance trait. Weed Res 43(6):404–417 (2003).
- 67 Maxwell BD, Roush ML and Radosevich SR, Prevention and management of herbicide resistant weeds, in *Proceedings of the 9th Australian Weeds Conference. Crop Science Society of South Australia*, Adelaide, SA, Australia, pp. 260–267 (1990).
- 68 Maxwell BD, Roush ML and Radosevich SR, Predicting the evolution and dynamics of herbicide resistance in weed populations. *Weed Technol* **4**(1):2–13 (1990).
- 69 Roush ML, Radosevich SR and Maxwell BD, Future outlook for herbicide-resistance research. Weed Technol 4:208–214 (1990).
- 70 Maxwell BD, Weed thresholds: the space component and considerations for herbicide resistance. Weed Technol 6(1):205 212 (1992).
- 71 Evans F, Diggle AJ and Renton M, Modelling the effects of farm management on the spread of herbicide resistance. Proc 21st Asian-Pacific Weed Science Society (APWSS) Conf, Colombo, Sri Lanka (2007).
- 72 Evans FH and Diggle AJ, A spatio-temporal modelling framework for assessing the impact of weed management technologies on the spread of herbicide resistance, in *Proceedings of the 16th Australian Weeds Conference*, ed. by van Klinken RD, Osten VA, Panetta FD and Scanlan JC. Queensland Weeds Society, Brisbane, Qld, Australia, pp. 218–220 (2008).
- 73 Richter O and Seppelt R, Flow of genetic information through agricultural ecosystems: a generic modelling framework with application to pesticide-resistance weeds and genetically modified crops. *Ecol Model* **174**(1–2):55–66 (2004).
- 74 Richter O, Zwerger P and Böttcher U, Modelling spatio-temporal dynamics of herbicide resistance. *Weed Res* **42**(1):52–64 (2002).
- 75 Richter O, Modelling dispersal of populations and genetic information by finite element methods. *Environ Model Software* 23(2):206–214 (2008).
- 76 Roux F, Paris M and Reboud X, Delaying weed adaptation to herbicide by environmental heterogeneity: a simulation approach. *Pest Manag Sci* 64(1):16–29 (2008).
- 77 Roux F and Reboud X, Herbicide resistance dynamics in a spatially heterogeneous environment. Crop Prot 26(3):335–341 (2007).
- 78 Baker J and Preston C, Predicting the spread of herbicide resistance in Australian canola fields. *Transgenic Res* **12**(6):731–737 (2003).
- 79 Colbach N, Clermont-Dauphin C and Meynard JM, GeneSys: a model of the influence of cropping system on gene escape from herbicide tolerant rapeseed crops to rape volunteers: II. Genetic exchanges among volunteer and cropped populations in a small region. Agric Ecosyst Environ 83(3):255 – 270 (2001).
- 80 Colbach N, Fargue A, Sausse C and Angevin F, Evaluation and use of a spatio-temporal model of cropping system effects on gene escape from transgenic oilseed rape varieties: example of the GeneSys model applied to three co-existing herbicide tolerance transgenes. Eur J Agron 22(4):417–440 (2005).
- 81 Putwain PD and Mortimer AM, The resistance of weeds to herbicides: rational approaches for containment of a growing problem. *Proc Brighton Crop Protection Conf Weeds*, BCPC, Farnham, Surrey, UK, pp. 285–294 (1989).



- 82 Warwick SI, Herbicide resistance in weedy plants: physiology and population biology. *Annu Rev Ecol Syst* **22**:95 114 (1991).
- 83 Groeters FR and Tabashnik BE, Roles of selection intensity, major genes, and minor genes in evolution of insecticide resistance. *J Econ Entomol* **93**(6):1580–1587 (2000).
- 84 Firbank LG and Watkinson AR, On the analysis of competition within two-species mixtures of plants. J Appl Ecol 22(2):503-517 (1985).
- 85 Firbank LG and Watkinson AR, A model of interference within plant monocultures. J Theor Biol 116(2):291 – 311 (1985).
- 86 Thornby D, Walker S and Whish J, Simulating weed persistence and herbicide resistance in the northern grain region using a validated crop growth model with extensions for seedbank dynamics and mating, in *Proceedings of the 15th Australian Weeds Conference*, ed. by Preston C, Watts JH and Crossman ND. Weed Management Society of South Australia, Adelaide, SA, Australia (2006).
- 87 Thornby DF and Walker SR, Simulating the evolution of glyphosate resistance in grains farming in northern Australia. *Ann Bot* **104**(4):747–756 (2009).
- 88 Neve P, Norsworthy JK, Smith KL and Zelaya IA, Modelling evolution and management of glyphosate resistance in *Amaranthus palmeri*. *Weed Res* **51**(2):99–112 (2011).
- 89 Neve P, Norsworthy JK, Smith KL and Zelaya IA, Modeling glyphosate resistance management strategies for Palmer amaranth (*Amaranthus palmeri*) in cotton. *Weed Technol* **25**(3):335–343 (2011).
- 90 Gressel J, Segel L and Ransom JK, Managing the delay of evolution of herbicide resistance in parasitic weeds. *Int J Pest Manag* **42**(2):113–129 (1996).
- 91 Bagavathiannan MV, Norsworthy JK, Smith KL and Neve P, Modeling the evolution of glyphosate resistance in barnyardgrass (*Echinochloa crus-galli*) in cotton-based production systems of the midsouthern United States. *Weed Technol* **27**(3):475–487 (2013).
- 92 Hanson DE, Ball DA and Mallory-Smith CA, Herbicide resistance in jointed goatgrass (Aegilops cylindrica): simulated responses to agronomic practices. Weed Technol 16(1):156–163 (2002).
- 93 Cavan G, Cussans J and Moss SR, Modelling different cultivation and herbicide strategies for their effect on herbicide resistance in Alopecurus myosuroides. Weed Res 40:561 – 568 (2000).
- 94 Cavan G, Cussans J and Moss SR, Managing the risks of herbicide resistance in wild oat. *Weed Sci* **49**:236–240 (2001).
- 95 Délye C, Jasieniuk M and Le Corre V, Deciphering the evolution of herbicide resistance in weeds. *Trends Genet* 23:1–4 (2013).
- 96 Delye C, Menchari Y, Guillemin JP, Matejicek A, Michel S, Camilleri C et al., Status of black grass (Alopecurus myosuroides) resistance to acetyl-coenzyme A carboxylase inhibitors in France. Weed Res 47(2):95–105 (2007).
- 97 Gressel J, Evolving understanding of the evolution of herbicide resistance. *Pest Manag Sci* **65**(11):1164–1173 (2009).
- 98 Shaw M, A model of the evolution of polygenically controlled fungicide resistance. *Plant Pathol* 38:44–55 (1989).
- 99 Renton M, The weeds fight back: individual-based simulation of evolution of polygenic resistance to herbicides, in *The 18th World IMACS Congress and MODSIM09 International Congress on Modelling and Simulation*. MSSANZ/IMACS, Cairns, Qld, Australia, pp. 574–580 (2009)
- 100 Langemann D, Richter O and Vollrath A, Multi-gene-loci inheritance in resistance modeling. *Math Biosci* 242(1):17–24 (2013).
- 101 Shi M, Collins PJ, Ridsdill-Smith TJ, Emery RN and Renton M, Dosage consistency is the key factor in avoiding evolution of resistance to phosphine and population increase in stored-grain pests. *Pest Manag Sci* 69(9):1049–1060 (2013).
- 102 Shi M, Renton M, Ridsdill-Smith J and Collins P, Constructing a new individual-based model of phosphine resistance in lesser grain borer (*Rhyzopertha dominica*): do we need to include two loci rather than one? *J Pest Sci* 85:1–18 (2012).
- 103 Gressel J, Molecular Biology of Weed Control. Taylor & Francis, London, UK (2002).
- 104 Renton M, Simulation modelling can help understand and predict how management, weed biology and genetics affect the development of herbicide resistance, in *Proceedings of the 24th Asian-Pacific Weed Science Society Conference*, ed. by Bakar B, Kurniadie D and Tjitrosoedirdjo S. Asian-Pacific Weed Science Society, Bandung, Indonesia, pp. 258–265 (2013).
- 105 Grimm V and Railsback SF, Individual-based Modeling and Ecology. Princeton University Press, Princeton, NJ (2004).
- 106 Renton M, Shifting focus from the population to the individual as a way forward in understanding, predicting and managing the

- complexities of evolution of resistance to pesticides. *Pest Manag Sci* **69**(2):171 175 (2012).
- 107 Uchmanski J and Grimm V, Individual-based modelling in ecology: what makes the difference? Trends Ecol Evol 11 (10):437 – 441 (1996).
- 108 DeAngelis DL and Mooij WM, Individual-based modeling of ecological and evolutionary processes. Annu Rev Ecol Evol Syst 36:147 – 168 (2005).
- 109 Breckling B, Middelhoff U and Reuter H, Individual-based models as tools for ecological theory and application: understanding the emergence of organisational properties in ecological systems. *Ecol Model* 194(1–3):102–113 (2006).
- 110 Gaba S, Cabaret J, Sauvé C, Cortet J and Silvestre A, Experimental and modeling approaches to evaluate different aspects of the efficacy of Targeted Selective Treatment of anthelmintics against sheep parasite nematodes. Vet Parasitol 171 (3-4):254-262 (2010).
- 111 Jaffe K, Issa S, Daniels E and Haile D, Dynamics of the emergence of genetic resistance to biocides among asexual and sexual organisms. J Theor Biol 188(3):289–299 (1997).
- 112 Vila-Aiub MM, Neve P and Powles SB, Fitness costs associated with evolved herbicide resistance alleles in plants. New Phytol 184(4):751–767 (2009).
- 113 Owen MJ, Michael PJ, Renton M, Steadman KJ and Powles SB, Towards large-scale prediction of *Lolium rigidum* emergence. II. Correlation between dormancy and herbicide resistance levels suggests an impact of cropping systems. *Weed Res* 51(2):133–141 (2011).
- 114 Vila-Aiub MM, Neve P, Steadman KJ and Powles SB, Ecological fitness of a multiple herbicide-resistant *Lolium rigidum* population: dynamics of seed germination and seedling emergence of resistant and susceptible phenotypes. *J Appl Ecol* 42(2):288 298 (2005).
- 115 Délye C, Menchari Y, Michel S, Cadet É and Corre VL, A new insight into arable weed adaptive evolution: mutations endowing herbicide resistance also affect germination dynamics and seedling emergence. Ann Bot 111(4):681 – 691 (2013).
- 116 Dyer WE, Chee PW and Fay PK, Rapid germination of sulfonylurea-resistant Kochia scoparia L. accessions is associated with elevated seed levels of branched chain amino acids. Weed Sci 41(1):18–22 (1993).
- 117 Hastings A, Cuddington K, Davies KF, Dugaw CJ, Elmendorf S, Freestone A et al., The spatial spread of invasions: new developments in theory and evidence. Ecol Lett 8(1):91 – 101 (2005).
- 118 Kuparinen A, Markkanen T, Riikonen H and Vesala T, Modeling air-mediated dispersal of spores, pollen and seeds in forested areas. *Ecol Model* **208**(2–4):177–188 (2007).
- 119 Renton M, Shackelford N and Standish RJ, Habitat restoration will help some functional plant types persist under climate change in fragmented landscapes. Global Change Biol 18(6):2057–2070 (2012).
- 120 Dauer JT, Luschei EC and Mortensen DA, Effects of landscape composition on spread of a herbicide-resistant weed. *Landsc Ecol* 24(6):735–747 (2009).
- 121 Bennett JC, Diggle A, Evans F and Renton M, Assessing eradication strategies for rain-splashed and wind-dispersed crop diseases. Pest Manag Sci 69(8):955–963 (2013).
- 122 Savage D, Barbetti MJ, MacLeod WJ, Salam MU and Renton M, Timing of propagule release significantly alters the deposition area of resulting aerial dispersal. *Divers Distrib* 16(2):288–299 (2010).
- 123 Busi R, Michel S, Powles SB and Délye C, Gene flow increases the initial frequency of herbicide resistance alleles in unselected *Lolium rigidum* populations. *Agric Ecosyst Environ* **142**(3–4):403–409 (2011).
- 124 Busi R, Yu Q, Barrett-Lennard R and Powles S, Long distance pollen-mediated flow of herbicide resistance genes in *Lolium* rigidum. Theor Appl Genet 117(8):1281–1290 (2008).
- 125 Dauer JT, Mortensen DA and Vangessel MJ, Temporal and spatial dynamics of long-distance Conyza canadensis seed dispersal. J Appl Ecol 44(1):105–114 (2007).
- 126 Délye C, Clément JAJ, Pernin F, Chauvel B and Le Corre V, High gene flow promotes the genetic homogeneity of arable weed populations at the landscape level. *Basic Appl Ecol* 11(6):504–512 (2010).
- 127 Sui R, Thomasson JA, Hanks J and Wooten J, Ground-based sensing system for weed mapping in cotton. Comput Electron Agric 60(1):31–38 (2008).
- 128 Cook T and Eldershaw V, Weed detecting technology: an excellent opportunity for advanced glyphosate resistance management, in Developing Solutions to Evolving Weed Problems – 18th Australasian



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- Weeds Conference, ed. by Eldershaw V. Weed Science Society of Victoria, Melbourne, Vic., Australia, pp. 245–247 (2012).
- 129 Andújar D, Escolà A, Dorado J and Fernández-Quintanilla C, Weed discrimination using ultrasonic sensors. Weed Res 51(6):543–547 (2011).
- 130 McCarthy C, Rees S and Baillie C, Machine vision-based weed spot spraying: a review and where next for sugarcane? *Proc Aust Soc Sugar Cane Technol* 32:424–432 (2010).
- 131 Beckie HJ, Hall LM and Schuba B, Patch management of herbicide-resistant wild oat (*Avena fatua*). *Weed Technol* **19**(3):697–705 (2005).
- 132 Pedersen SM, Fountas S, Have H and Blackmore BS, Agricultural robots system analysis and economic feasibility. *Precis Agric* **7**(4):295–308 (2006).
- 133 Michael PJ, Owen MJ and Powles SB, Herbicide-resistant weed seeds contaminate grain sown in the Western Australian grainbelt. Weed Sci 58(4):466–472 (2010).
- 134 Park SE, Benjamin LR and Watkinson AR, The theory and application of plant competition models: an agronomic perspective. *Ann Bot* 92(6):741–748 (2003).
- 135 Deen W, Cousens R, Warringa J, Bastiaans L, Carberry P, Rebel K et al., An evaluation of four crop:weed competition models using a common data set. Weed Res 43(2):116–129 (2003).
- 136 Berger U, Piou C, Schiffers K and Grimm V, Competition among plants: concepts, individual-based modelling approaches, and a proposal for a future research strategy. *Perspect Plant Ecol Evol Syst* 9(3–4):121–135 (2008).

- 137 Cici SZ-H, Adkins S and Hanan J, A canopy architectural model to study the competitive ability of chickpea with sowthistle. *Ann Bot* 101(9):1311–1318 (2008).
- 138 Dunbabin V, Simulating the role of rooting traits in crop-weed competition. *Field Crops Res* **104**(1–3):44–51 (2007).
- 139 Gaines TA, Zhang W, Wang D, Bukun B, Chisholm ST, Shaner DL et al., Gene amplification confers glyphosate resistance in Amaranthus palmeri. Proc Natl Acad Sci USA 107(3):1029–1034 (2010).
- 140 Basu C, Halfhill MD, Mueller TC and Stewart CN, Jr, Weed genomics: new tools to understand weed biology. *Trends Plant Sci* 9(8):391–398 (2004).
- 141 Malausa T, Gilles A, Meglécz E, Blanquart H, Duthoy S, Costedoat C et al., High-throughput microsatellite isolation through 454 GS-FLX Titanium pyrosequencing of enriched DNA libraries. Mol Ecol Resour 11(4):638–644 (2011).
- 142 Shi M, Collins PJ, Ridsdill-Smith J and Renton M, Individual-based modelling of the efficacy of fumigation tactics to control lesser grain borer (*Rhyzopertha dominica*) in stored grain. *J Stored Prod Res* **51**:23–32 (2012).
- 143 Gressel J, Low pesticide rates may hasten the evolution of resistance by increasing mutation frequencies. *Pest Manag Sci* 67(3):253–257 (2011).
- 144 Peck SL, Antibiotic and insecticide resistance modeling is it time to start talking? *Trends Microbiol* **9**(6):286–292 (2001).
- 145 REX Consortium, Structure of the scientific community modelling the evolution of resistance. PLoS ONE 2(12):e1275 (2007).